Senecio vulgaris L. (Asteraceae) **Common Groundsel**

Description. Annual, herbaceous, from a thin, branched taproot; stems 8-60 cm tall, erect to decumbent, somewhat succulent, sparsely tomentose when young, glabrous to sparsely villous, simple or branched near the base. Leaves 2-5(10) cm long, 0.5-4.5 cm wide, deeply pinnately lobed or toothed, glabrous to sparsely villous, the lobes oblong, irregularly toothed, the lower ones petioled, oblanceolate to obovate, base abruptly tapered, the upper ones oblong, sessile or clasping, the bases sometimes auriculate. Heads discoid (all corollas radial and salverform), 5-10 mm long, 4-10 mm wide, cylindrical to subcylindrical, subsessile to stalked, 8-20, in terminal or axillary clusters. Phyllaries in two series, glabrous to sparsely pubescent, green, black-tipped, the outer ones 2-10, 1-2 mm long, the inner ones 21, 4-6 mm long, linear-lanceolate, margins scarious, apices acute, glabrous to loosely tomentose. Corollas yellow. Achenes 1.5-2 mm long, brown, glabrous or with appressed-puberulent ribs; pappus 15-20 mm long, simple, white. In California, flowering from January through December. (Barkley 1993, Chater and Walters 1976, Clapham et al. 1962, Cronquist 1980, 1994, Fernald 1950, Great Plains Flora Association 1986, Munz 1959, Welsh et al. 1987).

Senecio vulgaris is a polyploid (2n=40), believed to have been derived through hybridization involving at least one extant species, S. squalidus L. (Ashton and Abbott 1992, Harris 1992, Stace 1977). Introgressive hybridization from S. squalidus has contributed to genetic variability in European populations (Abbott et al. 1992), including forms with ray flowers (Abbott et al. 1990, Ingram and Noltie 1984, Ingram et al. 1980). Such races are not apparently present in introduced populations, at least in California as inferred from descriptions in Barkley (1993) and Munz (1959). Naturalized British populations show considerable genetic variation (Theaker and Briggs 1992, 1993). Genetic variation included the evolution of strains with varying growth rates, this in response to natural selection imposed by weed removal methods (Kadereit and Briggs 1985, Theaker and Briggs 1993).

Geographic distribution. Native and widespread throughout continental Europe, but more common in the Mediterranean region. It has been introduced into Australia, New Zealand, throughout North America, Great Britain [repeatedley according to Stace (1997) and Ingram et al. 1980)], Japan, southern Africa (Arnold and de Wet 1993, Chater and Walters 1976, Clapham et al. 1962, Cronquist 1980, 1994, Fernald 1950, Ohwi 1953, Webb et al. 1988).

Common groundsel was first reported from California ("near San Francisco") in 1876 (Brewer et al. 1876). Naturalized populations occur on all the Channel Islands except for Santa Barbara (Junak et al. 1995), coastal California from Del Norte County southward to San Diego County, and in most counties west of the Sierra Nevada (Anonymous 1998, Barkley 1993).

Ecological distribution. In its natural range, common groundsel occurs in cultivated and fallow fields (Chater and Walters 1976, Clapham et al. 1962). It has been reported elsewhere from similar habitats (e.g., (Barkley 1993, Fernald 1950, Munz 1959, Welsh et al. 1987), in turf (Murphy 1996), and in household gardens (Senesac 1991).

Reproductive and vegetative biology. Common groundsel is self-compatible, but experiences some outcrossing in native populations (Abbott et al. 1990, Marshall and Abbott 1984a, 1984b). Reproductive capacity is high relative to total plant biomass (Harper and Ogden 1970). Small, light seeds and proportionately large pappus confer a relatively high level of dispersability (Anderson 1992, Sheldon and Burrows 1973)

In Britain, common groundsel seeds may germinate at any time of the year, depending on availability of water (Harper and Ogden 1970). Most plants live for about 3 months; flowering and reproduction begins within 6-8 weeks. A comparison of strains from wild Scotish and Spanish populations demonstrated differences in the proportion of dormant seeds and rates of germination, which were related to differences between cold- and warm-temperate (Mediterranean) climates (Ren and Abbott 1991). In general, seeds of cold-temperate strains do not germinate unless they have been exposed to freezing temperatures; those of warm-temperate (Mediterranean) climates display little dormancy, germinating only after sufficient precipitation. However, buried seeds may remain viable and dormant for as long as 5 years (Roberts 1964).

Using experimental populations differing in patchiness and density, Bergelson (1990) demonstrated that establishment and reproductive capacity was greater when competing species (i.e., Poa annua) were clumped rather than randomly distributed; this suggests that common groundsel may not be competitive under conditions of relatively dense cover (Bergelson et al. 1993). This is consistent with observations that common groundsel is invasive and common in disturbed habitats (Harper and Ogden 1970, Murphy 1996, Senesac 1991), and does not compete well under horticultural practices that minimize open space (Qasem and Hill 1993). Common groundsel also has been reported as invasive after fires, both in European shrubland (Trabaud 1991) and in California chasparral (Sweeney 1956, Biswell 1974). However, in both instances, it does not persist except in disturbed sites.

Allelopathy (Oasem and Hill 1989), nitrogen levels (Oasem and Hill 1993), and both intraand inter-specific interactions among seedlings (Bergelson and Perry 1989) have been implicated as factors conferring competitive advantage to persistence in naturalized populations.

Weed status. Common groundsel is not considered a serious noxious weed in agricultural or horticultural practice, at least at a global level (not listed by Holm et al. 1977), nor is it considered a noxious weed by the State Dept. of Food and Agriculture (Anonymous 1996). However, it has been reported as weedy and invasive, primarily in disturbed sites, fallow or cultivated (both agricultural and horticultural) fields, in turf, and in urban gardens, where it is able to germinate and grow at any time the year (Murphy 1996, Senesac 1991).

Microbial pathogens. Several studies have investigated natural fungal pathogens (Ayres 1995, Baka and Losel 1992, Hallett et al. 1990a, 1990b, Hallett and Ayres 1992, Harry and Clarke 1992, Paul 1989, Paul and Ayers 1987b, 1990, Preese 1987), which include mildews (Albugo spp., Botrytis spp., Erysiphe fischeri, E. cichoracearum, Sphaerotheca epilobii) and rusts (Puccinia lagenophorae, P. punctiformis, Uromyces spp.). Most of these studies, however, are based on greenhouse conditions or in horticultural (e.g., flowers) and agricultural production (e.g., lettuce).

Application of fungal diseases to biocontrol of *Senecio vulgaris* has been discussed by Paul et al. (1993). Damage by *Puccinia lagenophorae* is enhanced by mild drought conditions, periods of frost in winter, and by competition between groundsel and neighbouring plants, but is reduced

by nutrient deficiency. Rust injury also is greatly increased by secondary infection of pustules by necrotrophic fungi. Other interactions that influence survival include the extent of mycorrhizal infection and phosphate levels in the soil (West 1995a, West 1995b), availability and composition of nutrients (Paul and Ayres 1986a, Paul and Ayres 1990), season of infestation (Paul and Ayres 1986c, Paul and Ayres 1986d, Paul and Ayres 1986e, Paul and Ayres 1987a), and intra- or inter-specific competition (Paul and Avres 1986b, Paul and Avres 1987b, Paul and Ayres 1990). A few studies have shown that *Senecio vulgaris* can evolve fungal-resistant strains (Harry and Clarke 1986, 1987).

Insect pathogens. No literature that reported common groundsel as a host of insect pathogens was found. Common groundsel produces pyrrolizidine alkaloids (e.g., Hartmann and M. Zimmer, 1986. Pieters and Vlietinck 1988), which are known to be either toxic (McHenry et al. 1990, Mitich 1995) or distasteful (Blaney and Simmonds 1985) to potential herbivores.

Herbicide control. The effectiveness of over 10 different herbicides (e.g., metolachlor, acifluorfen, simazine) on groundsel were tested (Al-Khatib 1995, Gallitano and Skroch 1993, Sym 1988), but primarily in association with agricultural crops or container propagation. Several studies studied various levels of resistance to such herbicides as atrazine, simazine, and triazine (Delaney 1984, Derr 1991, Fuerst et al. 1986, Havaux 1989, McCloskey and Holt 1990, 1991, Stephenson et al. 1990, Watson et al. 1987). Watson et al. (1987) provided evidence for relatively simple inheritance (a few loci) for herbicide resistance. Stowe and Holt (1988) examined physological mechanisms that confer herbicide resistance by comparing triazineresistant, non-resistant, and hybrid strains. Holt and Goffner (1985) studied morphological and physiological traits associated with simazine resistance.

Relatively rapid selection for genetic strains differing in cultural conditions, minor morphological traits, and herbicide resistance has been demonstrated in some populations (Briggs and Block 1992, Briggs et al. 1992, Radosevich 1973). Holliday and Putwain (1977, 1980) showed that recruitment of simazine-resistant strains came from resident seed banks, rather than from immigration seeds, because resident strains germinated, survived, and reproduced during times coincident with low herbicide levels.

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